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**IgM Capture Bestarose HP
Affinity
chromatography resin
Instruction for use**



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1. Introduction

IgM Capture Bestarose HP is a thiophilic affinity resin made by fixing the sulfur-containing compound 2-mercaptopyridine on fine particles of high rigidity agarose. Its optimized ligand density has an appropriate affinity with IgM while its fine particles can increase the binding capacity of IgM with larger molecular weight. Thiophilic affinity works by the interaction between electron donor and electron acceptor to isolate and purify biomolecules, which is strengthened in high-salt environments and weakened in low-salt environments

2. Technical characteristics

Appearance	White slurry
Matrix	Highly cross-linked agarose, 6%
Particle size	24~44 μ m (the average particle size is 34 μ m)
Functional group	2-mercaptopyridine
Ligand concentration	~2mg 2- mercaptopyridine /mL resin
Dynamic binding capacity	5mg human IgM/mL resin
Chemical Stability	Stable in common aqueous buffers:30% isopropyl alcohol,70% ethanol,1M HAC+,0.1M NaOH
Max. pressure	0.3MPa
pH stability	2~13(CIP),3~11(Working)
Storage++	2~30 $^{\circ}$ C, 20% ethanol or 2% benzyl alcohol

+ 1M HAC only be used for cleaning.

++2% benzyl alcohol is only used for international transport or special requirements from customer.

3. Method of chromatography

3.1 Column packing

Note: It is best to equilibrate the resin slurry to room temperature before column packing.

Due to the fine particle size of the resin, a chromatography column with a mesh size of 10 microns or less should be selected.

- According the column volume to calculate the amount of resin.

Resin volume=column volume \times 1.15(Compression factor=1.15)

According to the volume of the suspended resin required, the suspended slurry of the resin required is calculated by the follow:

Required resin slurry¹ volume = Suspended resin volume \div Resin slurry¹ concentration.The original concentration of resin slurry¹ is shown in the follow table.



Pack size	Resin slurry ¹ concentration (%)
25mL,100mL,500mL,1L,5L,10L	80
20L,40L	75

1: It refers to the original packaging resin slurry sold by Bestchrom.

Note: For resins in non-original concentration, customer can calculate the required volume according to the actual concentration of resin slurry.

- Washing the resin: Suspend the resin by shaking and pour into a funnel, remove the liquid, and wash with about 3mL packing solution (0.1M NaH₂PO₄ with 1.2M (NH₄)₂SO₄, pH7.0)/mL resin for 3 times. Use a glass stick or stirrer to stir each time you add the packing solution, in order to better clean the shipping buffer.
- Prepare the packing slurry: Transfer the washed resin from the funnel into a beaker or other appropriate container, add packing solution to obtain a 50%~75% slurry, stir well and set aside for use.
- Take a cleaned B XK column (B XK series columns with diameters ranging from 1cm to 30cm can satisfy different scale chromatography applications). Take B XK16/20 for example, purge the bubbles trapped at the end-piece net by draining some packing solution through the column outlet. Leave about 1cm water at the bottom of the column and close the bottom outlet. Adjust the column so that it is perpendicular to the ground.
- Slowly pour the slurry into the column at one time (use a packing reservoir if necessary). Do not bring any air bubbles into the column.

Packing reservoir: Empty glasstube with same diameter as the B XK column.

- Fill the remainder of the column with packing solution. Connect the packing reservoir to the chromatography system, open the flow velocity, drain the bubbles in the hose, close the flow velocity, and tighten the top cover of the packing reservoir.
- ◇ After pouring, stir well again with Stirrer, and then wash the resin particles on the inner wall of the column from top to bottom with the packing solution, and let the resin settle naturally until there is about 1cm of clarifying solution on the suspension. Mount the adapter and connect the adapter to the chromatography system or peristaltic pump. Lower the adapter to descend to contact with the clarifying solution and tighten the sealing ring after it is fully immersed in the clarifying solution. With the outlet of the top piece is opened, slowly move the adapter down until all bubbles are drained.

Note: This operation is only applicable to B XK 100 and above columns. Flushing the inner wall reduces the resin particles sticking between the seal ring and the column wall, avoiding the risk of leakage.

- When the bed height is 10cm, the flow velocity can be set to 250cm/h. Open the bottom plug, start the pump and run the setting flow velocity until the bed is stabilized, mark the bed height.
- Remove the packing reservoir (if any), mount the adaptor, lower the adaptor to about 0.5cm above the resin surface, and continue to press the column using the above flow velocity until the bed is completely consolidated, mark the consolidated bed height.



- Stop the pump, open top plug, close the bottom plug, loosen the O-ring seal slightly, press the adaptor to about 0.3cm below the marked position, tighten the O-ring seal, close adaptor stop plug, and complete the column packing.

3.2 Evaluation of Packing

- The packing quality of chromatographic column can be confirmed by column efficiency measurement and evaluation. The tests are required after the column packing, during the column working life and when the separation and purification performance weakens. The method usually relies on the height equivalent to a theoretical plate (HETP) and the asymmetry factor(As).
- Acetone or NaCl solution can be used as sample for the testing. Sample solution and mobile phase can be prepared according to the following table.

	Acetone method	NaCl method
Sample	1.0% (v/v)acetone in water	0.8M NaCl in water
Sample volume	1.0%CV	1.0%CV
Mobile phase	Water	0.4M NaCl in water
Flow velocity	30cm/h	30cm/h
Detection	UV280 nm	Conductivity

- Method for measuring HETP and As:

Use UV curve or the conductivity curve to calculate the height equivalent of theoretical plate (HETP), number of theoretical plates(N) and the asymmetry (As):

$$HETP=L/N$$

$$N=5.54(V_R/W_h)^2$$

Note: V_R = retention volume

W_h = half-peak width

L = column height

N = the number of theoretical plates

(The units of V_R and W_h should be the same)

$$As=b/a$$

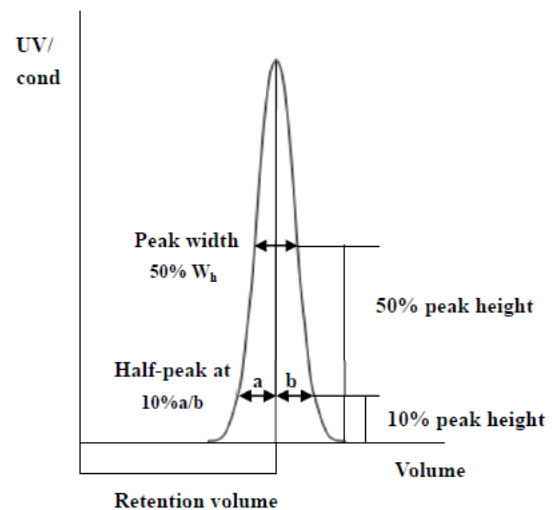
Note:

a= 1st half peak width at 10% of peak height

b= 2nd half peak width at 10% of peak height

- Evaluation the column packing

As a guideline, if the value of HETP is less than 3 times the average particle size(d_{50}) of the resin and the As is between 0.8~1.8, the column is very efficient. The unsatisfactory results should be analyzed and the column should be repacked .



3.3 Chromatographic method

- Recommended buffer:
Binding buffer: 20mM PB, 0.8M $(\text{NH}_4)_2\text{SO}_4$, pH7.59 (some monoclonal IgM may not bind to the column in 0.8M $(\text{NH}_4)_2\text{SO}_4$, and the concentration of $(\text{NH}_4)_2\text{SO}_4$ can be increased to 1M to improve the binding ability)
Elution buffer: 20mM PB, pH7.5
Cleaning buffer: 20mM PB, 30% isopropyl alcohol, pH7.5
In some cases, 0.8M ammonium sulfate can be replaced by 0.5M potassium sulfate. Most monoclonal IgM can bind to a column at 0.5M potassium sulfate, and the purity of monoclonal IgM purified from 0.8M ammonium sulfate and 0.5M potassium sulfate is comparable.
- Flow velocity: According to the column bed height to set the flow velocity (generally less than 150cm/h), the higher column bed height is, the lower flow velocity will be.
- Sample preparation: In order to prevent the column blocking, before the loading sample, it needs to be filtered by 0.45 μm microporous membrane, and the pH and conductivity of the sample should be adjusted to same as equilibration buffer. The concentration of $(\text{NH}_4)_2\text{SO}_4$ affects the binding capacity.
- Equilibration: Washing the column with equilibration buffer, which usually needs 3-5CV.
- Loading sample: The loading volume is determined by the substance content in the sample and the binding capacity of IgM Capture Bestarose HP.
- Cleaning: Wash the column with equilibration buffer until the UV absorption value is close to baseline.
- Elution: Elution peaks can be collected using the recommended elution buffer. If the target protein elution is incomplete, 20mM PB pH7.5 buffer containing a low concentration (e.g., 5-10%) of isopropyl alcohol may be used, but high concentration of isopropyl alcohol should not be used
- Regeneration: Rinse the column with cleaning buffer.
- Re-equilibration: After rinsing with equilibration buffer, the second sample can be loaded, repeat the process if necessary.

4. Cleaning-in-place (CIP)

With the increasing use of chromatography resin, the accumulation of contaminants on the chromatography column is also increasing. Cleaning-in-place can prevent the accumulation of contaminants and maintain a stable working state. Determine the frequency of CIP according to the degree of contamination of the resin (if the contamination is serious, CIP should be carried out after each use to ensure repeatability of the results).

- Modified protein: Wash with 4CV of 0.1M NaOH, then rinse immediately with 5CV of equilibration buffer.
- Strongly hydrophobic substance or lipid: Wash with 2~4CV of 20mM PB, 30% isopropyl

alcohol, pH7.5 buffer. Then flush immediately with equilibration buffer of at least 5CV.

5. Sterilization

Since the 20% ethanol or 2% benzyl alcohol preservation solution does not have sterilization and depyrogenation, it is recommended that IgM Capture Bestarose HP can be treated with 70% ethanol for more than 12h to achieve the purpose of sterilization and removal of pyrogens.

6. Storage

IgM Capture Bestarose HP is supplied in 20% ethanol or 2% benzyl alcohol. It should be stored in 20% ethanol and sealed at 2-30°C after use. In order to prevent ethanol volatilization and microbial growth, it is recommended to replace the storage solution regularly.

7. Disposal and recycling

IgM Capture Bestarose HP is very difficult to degrade in nature, incineration is recommended to protect the environment.

8. Order information

Product	Cat. No.	Pack size
IgM Capture Bestarose HP	AA0152	25mL
	AA0153	100mL
	AA0154	500mL
	AA0155	1L
	AA0156	5L
	AA0157	10L

Prepacked columns	Cat. No.	Pack size
EzFast IgM HP	EA01521	1×1mL
	EA01531	5×1mL
	EA01523	1×5mL
	EA01533	5×5mL
EzScreen IgM HP	EA01525	1×4.9mL
	EA01535	5×4.9mL
EzLoad 16/10 IgM HP	EA01501	1 pcs
EzLoad 26/10 IgM HP	EA01511	1 pcs